

Table 1

Examples of Autoimmune Diseases or Disease Model Caused
By Autoreactive B Cell Responses

Disease	Pathogenic Antibody Specificity
Myasthenia Gravis (MG)	Anti-acetylcholine receptor antibodies cause weakness in MG
Juvenile Onset Diabetes Mellitus (Type 1 Diabetes)	Anti-insulin antibodies and anti-islet cell antibodies mediate islet cell destruction
Graves' Disease	Anti-thyroid stimulating hormone receptor antibodies mediate the disease
Insulin Resistance in Diabetes Mellitus	Anti-insulin antibodies prevent treatment of diabetes with insulin

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Table 2

Examples Autoimmune Diseases or Disease Models Caused
by Autoreactive T Cell Responses

Experimental autoimmune uveoretinitis (EDU)	T cell responses against retinal S antigen cause eye damage
Experimental autoimmune encephalomyelitis (EAE)	T cell responses against myelin basic protein cause neuronal damage

005590-09101
TOT250-045550

Table 3

Peptide Sequences Used In Chimpanzee Immunizations

F-T1-SP10IIIIB(A)	AVGIGALFLGLKQIINMWQEVGKAMYACTRPNNNTRKSIRIQRGPGRAFVTI
T1-SP10IIIIB	KQIINMWQEVGKAMYACTRPNNNTRKSIRIQRGPG
T1SP10IIIIB(A)	KQIINMWQEVGKAMYACTRPNNNTRKSIRIQRGPGRAFVTI

Table 4

Tritiated Thymidine Incorporation of Peripheral Blood Mononuclear Cells Following In Vitro Stimulation With HIV Env gp120*

Chimpanzee No.	Immunogen	Pre-	Post-
		Immunization Δcpm/10 ⁶ cells (Post/Pre)	Immunization
884	T1-SP10IIIB, then T1-SP10IIIB(A)	169	39,189 (232)
1028	T1-SP10IIIB, then T1-SP10IIIB(A)	17,955	129,121 (7)
1045	F-T1-SP10IIIB(A)	6,348	12,256 (2)
1070	F-T1-SP10IIIB(A)	11,285	22,719 (2)

*Data represent the peak gp120 responses observed during the immunization period. Data for animals 884, 1028, and 1045 represent peak responses using from 2ug/ml to 0.5ug/ml of HIVIIIB(LAI) recombinant gp120. Data for animal 1070 represent peak responses using from 1ug/ml to 0.5ug/ml of native HIVIIIB(LAI) gp120.

Table 5

HIV Envelope gp41 Fusion Protein (F) Sequences From Multiple
HIV Isolates

Isolate	Sequence
<hr/> HIV-1	
BH10	A V G : I G A L F L G F L
MN	A A : : - - - - -
SC	- - - T - - - M - - - - -
SF2	- - - I V - - - M - - - - -
CDC4	- - - M L - - - M - - - - -
WMJ2	- - - T - - - M - - - - -
RF	- - - T - - - M - - - - -
ELI	- I - : L - - M - - - - -
MAL	- I - : L - - M - - - - -
Z6	- I - : L - - M - - - - -
Z321	- I - M : - - F - - - - -
JY1	- I - : L - - V - - - - -
WMJ-1	- - - A - - - M - - - - -
 HIV-2	
ROD	R G V F V L G F L G F L
NIHZ	- - - - -

Sequences for BH10 are aa 519-530 from Ratner, L, et al. Nature 313: 277-284, 1985. Sequences for the remainder of the HIV-1 and HIV-2 isolates from Myers, et al. Human Retroviruses and AIDS, 1988, Los Alamos National Laboratory, Los Alamos, New Mexico, p. II-90. WMJ-1 sequence from ref. 18.

Table 6

Regions of the TSH Receptor to Which Patient
Anti-TSH Receptor Autoantibodies Bind

Amino Acid No.	Sequence	Ref.
333-343	YVFFEEQEDEI	17
12-36	HQEEDFRVTCKDIQRIPSLPPSTQT	18
289-317	LRQRKSVNALNSPLHQEYEENLGDSTVGY	18
352-366	YVFFEEQEDEIIGF	27
103-111	YKELPLLKFL	28

Amino acid numbers and sequence from the reference listed

T0260-04695650

Table 7

Examples of Hybrid Peptide Constructs That Could Be Used To
Treat Anti-HLA Immune Responses In AIDS

HIV gp120 homology with DP/DQ β chain
gp120 aa261-270 VVSTQLLLNG
HLA DP/DQ aa142-151 VVST*LI*NG

HIV gp41 homology with HLA DR β chain
gp41 aa837-844 EGTDRVI
HLA DR aa19-25 NGTERVR

Hybrid Immunogens: AVGIGALFLGFLVVSTQLLLNG
 AVGIGALFLGFLVVSTLING
 AVGIGALFLGFLEGTDRVI
 AVGIGALFLGFLNGTERVR

HIV gp120 and gp41 homologies with HLA Class II are from refs. 25
and 26.

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TABLE 8

Sequences of Synthetic Peptide Constructs
Derived From HIV MN and HIVIIB Env gp120*

Peptide Name	Peptide Type	Peptide Composition and Sequence (Epitope Type)		
		F	T1 (Th)	SP10 (B cell)
F-T1-SP10IIB(A)	F-Th-B	AVGIGALFLGLKQIINMWQEVGKAMYACTRPNNTNRKSIRIQRGPGRFVVTI		A(B cell)
T1-SP10IIB(A)	Th-B	KQIINMWQEVGKAMYACTRPNNTNRKSIRIQRGPGRFVVTI		
T1-SP10IIB	Th-B	KQIINMWQEVGKAMYACTRPNNTNRKSIRIQRGP		
T1-SP10MN(A)	Th-B	KQIINMWQEVGKAMYACTRPNYNNKRKRIRIHIGPGRFYTTK		

*Each amino acid is represented by a single-letter code that is the first letter of its name, except for arginine (R), asparagine (N), glutamine (Q), glutamic acid (E), lysine (K), phenylalanine (F), tryptophan (W), tyrosine (Y), and aspartic acid (D). F (fusogenic domain) sequence is amino acids 519-530 from HIVIIB (27). T1 sequence is amino acids 428-443 from HIVIIB (27). SP10MN(A) sequence is amino acids 301-319 from HIVMN (28). SP10IIB sequence is amino acids 303-321 from HIVIIB. (A) sequence is amino acids 320-324 from HIVMN (28) and amino acids 322-327 from HIVIIB (27).

Th = T helper cell determinant.
B cell = B cell neutralizing antibody determinant.
A = Additional HIV gp120 V3 loop sequences added to the original synthetic peptide (SP10) sequence to add an additional neutralizing and CTL region to the HIV B cell determinant of the hybrid peptide.

27 = Ratner et al, Nature 313:277 (1985)
28 = Myers et al, Human Retroviruses and AIDS (1991), p. III 6-23

TABLE 6

Time Course of Anti-Peptide Antibody Responses in Chimpanzees
Immunized with HIV Envelope Synthetic Th-B or F-Th-B Peptides

Month of Study	Immunogen	Chimpanzee Number		Immunogen	Chimpanzee Number	
		884	1028		1045	1070
		Reciprocal of ELISA Titer			Reciprocal of ELISA Titer	
1		0	0		0	0
2	Th-B(IIIB) 6mg	0	0	F-Th-B(IIIB) 6mg	0	0
3	Th-B(IIIB) 6mg	51,200	102,400	F-Th-B(IIIB) 6mg	0	0
4	Th-B(IIIB) 6mg	25,600	819,200	F-Th-B(IIIB) 6mg	0	800
5	Th-B(IIIB) 6mg	25,600	204,800*	F-Th-B(IIIB) 6mg	1,600	200
6	Th-B(IIIB) 30mg	51,200	102,400	F-Th-B(IIIB) 30mg	25,600	12,800
7	Th-B(IIIB) 30mg	204,800	102,400	F-Th-B(IIIB) 30mg	25,600	12,800
8	Th-B(IIIB) 30mg	51,200	25,600	F-Th-B(IIIB) 30mg	6,400	12,800
9		51,200	51,200		3,200	6,400
10		12,800	25,600		800	800
11		51,200	25,600		800	1,600
12		51,200	25,600		1,600	800
13		25,600	25,600		200	200
14	Th-B(IIIB) 6mg	51,200	25,600	F-Th-B(IIIB) 1mg	200	400
15		102,400	12,800		800	800
16	Th-B(MN) 6mg	25,600	12,800	Th-B(IIIB) 6mg	100	0
17	Th-B(MN) 6mg	12,800	3,200	Th-B(MN) 6mg	1,600	3,200
18		25,600	6,400		6,400	25,600
19	Th-B(MN) 6mg	25,600	1,600	Th-B(MN) 6mg	6,400	51,200
20		51,200	6,400	Th-B(MN) 6mg	51,200	102,400#

Titers are endpoint ELISA titers (titers at which E/C were ≥ 3.0) against the Th-B peptide, Tl-SP10IIIB. * Animal 1028 did not receive the month 5 injection due to a sterile abscess at the injection site. All injections in animal 1028 after month 5 were in PBS alone.

Animal 1070 did not receive the month 20 immunization due to the presence of high levels of anti-HIV neutralize antibodies.

For animals 884 and 1028, immunizations at months 2-5 were with Tl-SP10IIIB, months 6,7,8 and 14, Tl-SP10IIIB(A). For animals 1045 and 1070 immunization at month 16 was with Tl-SP10IIIB(A).

Table 40
Mean Lymphocyte and Lymphocyte Subset Levels in Chimpanzees
Before and During Immunization With HIV Envelope Synthetic Peptides*

Lymphocyte Subset	Chimpanzee Number											
	884			1028			1045			1070		
	Before	During	% Change	Before	During	% Change	Before	During	% Change	Before	During	% Change
	Cells/mm ³ ± SEM											
Total	4034±452	3046±249	-26%	3164±396	3286±660	+4%	3164±397	1426±116	-55%	3943±885	2768±296	-30%
Lymphocytes	2629±384	2054±178	-24%	2565±276	2027±402	-21%	2460±253	1012±82	-59%	3337±762	1887±184	-44%
T cells	356±47	365±39	+3%	411±103	458±47	+11%	293±32	175±15	-40%	302±53	232±22	-23%
B cells	345±82	317±43	-9%	257±25	434±128	+68%	112±27	61±7	-45%	478±148	306±44	-36%
NK cells												

*Before samples were studied over a 5 month period prior to immunization with peptides; n = 5 for lymphocytes, n = 3 for T cells, B cells and NK cells. "During" samples were taken from months 2-14 of immunization; n = 11 for lymphocytes, T, B, and NK cells. Unless noted, p values for percent change comparing "before" values with "during" values was not significant with p > .05 using student's t test.

† = p > .001
‡ = p > .02
§ = p > .005

Table 11

Neutralization of HIV LAI/IIIB and HIV MN in Syncytium Inhibition Assay in Chimpanzees Immunized with T1-SP10 Peptides

Animal No.	Month 18		Month 19		Month 20	
	LAI/IIIB	MN	LAI/IIIB	MN	LAI/IIIB	MN
	Presence of Neutralization in Syncytium Inhibition Assay					
	(Reciprocal Titer in RT Inhibition Assay)					
884	-	- (20)	-	-	-	- (24)
1028	-	-	-	-	-	-
1045	-	- (23)	+/- (23)	- (23)	- (22)	- (24)
1070	+/- (92)	- (22)	+ (100)	+ (96)	+/- (86)	++ (350)

- = < 48% inhibition of syncytia.
 +/- = ≥ 49% and < 80% inhibition of syncytia.
 + = ≥ 80% inhibition of syncytia, titer 1:10.
 ++ = ≥ 80% inhibition of syncytia, titer 1:20.

Table 12

Reactivity of Chimpanzee Serum with Truncated Forms of
the Th-B Peptide T1-SP10IIIB[#]

Chimpanzee No. (Bleed Date)	Peptide Used in ELISA Binding Assay				
	T1-SP10IIIB	T1-flu	SP10C	SP10D	SP10E
	Endpoint Titer (> 3.0 E/C) In ELISA Assay				
884 (Month 7)	204,800	800	> 102,400*	51,200	3,200
1028 (Month 7)	102,400	800	102,400	51,200	3,200

[#]Peptides used in ELISA Assay were:

T1-SP10IIIB	-	KQIINMWQEVGKAMYACTRPNNNTRKSIRIQRGPG
T1-flu	-	KQIINMWQEVGKAMYATYQRTALVTG
SP10C	-	(C)TRKSIRIQRGPGR(Y)
SP10D	-	(C)IRIQRGPGR
SP10E	-	(C)TRPNNNTRKSIR

ELISA assay performed as described in Methods.

Flu sequence (TYQRTALVTG) is from influenza nucleoprotein, strain A PR/8/34 from Deres et al, Nature 342:561 (1989).

at 1:102,400 = 6.0.

Table 13

Effect of Derivatizing T1-SP10IIIIB(A) Peptide With
the HIV gp41 Fusogenic (F) Domain on Peptide
Ability to Bind to Human Cells .

Peptide	Antibody	MFC 4 Degrees C, 1 Hr.	MFC 37 Degrees C, 21 Hr.
None	Anti-gp120	7.6	13.6
T1-SP10IIIIB(A) 10ug/ml	Anti-gp120	14.7	14.0
F1-T1-SP10IIIIB(A)	Anti-gp120	82.8	36.7

Anti-gp120 monoclonal antibody was 0.5beta from the NIAID AIDS Research and Reference Reagent Program (Matsushita et al J. Virol. 62:2107, 1988). Cells used were human JY B cells which were incubated either for 1 hour at 4 degrees C or for 21 hours at 37 degrees C and then reacted with saturating amounts of the anti-gp120IIIIB mab, 0.5beta followed by FITC-conjugated goat anti-mouse Ig reagent. The amount of fluorescence was determined on a flow cytometer and fluorescence brightness was expressed as MFC=mean channel fluorescence.

Table shows that conjugation of the F domain on the T1-SP10IIIIB(A) peptide confers on it the ability to bind to JY B cells better than the T1-SP10IIIIB(A) peptide alone, and that after incubation at 37 degrees C, the F-T1-SP10IIIIB(A) peptide is decreased on the surface of the cells.

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Table 14

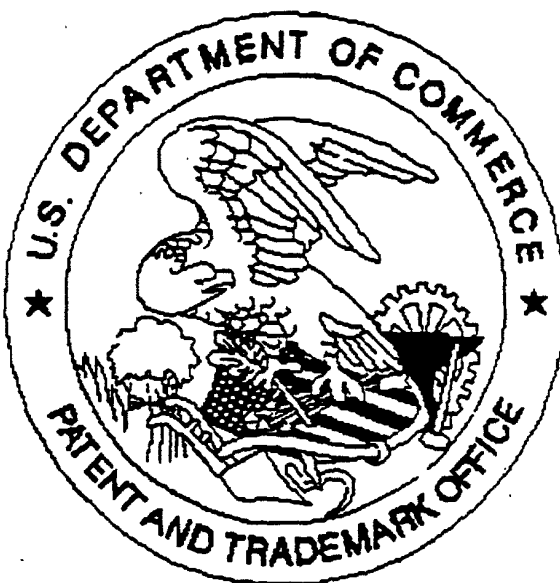
Reactivity of anti-gp120 Monoclonal Antibody with
Acetone-Fixed JY B Cells That Had Been Incubated
With F-T1-SP10IIIB(A) Peptide (10 μ g/ml) For 21
Hours at 37 Degrees C

Peptide	Antibody	% Intracytoplasmic Positive
T1-SP10IIIB(A)	Control	0
T1-SP10IIIB(A)	Anti-gp120	0
F-T1-SP10IIIB(A)	Control	0
F-T1-Sp10IIIB(A)	Anti-gp120	76 faint, 24 bright

Cells were incubated as described in Table 13.
After 21 hours at 37 degrees C, cytocentrifuge
preparations of cells were prepared, acetone
fixed, and reacted either with control mab P3x63
Ag8 or with anti-gp120 mab 0.5beta. Slides were
read for either faint or bright intracytoplasmic
fluoresence on a fluorescence microscope. Data
show that after incubation of 10 ug/ml of peptide
for 21 hours at 37 degrees C, the F-T1-SP10IIIB(A)
peptide could be detected inside the JY B cells
whereas the T1-SP10MN(A) peptide could not be
detected.

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